



Contribution of Catalase Positive Cocci on Flavour Formation in Fermented Sausages

Aybike Kamiloğlu^{1*}, Güzin Kaban² and Mükerrerem Kaya²

¹Department of Food Engineering, Faculty of Engineering, Bayburt University, 69000, Bayburt, Turkey.

²Department of Food Engineering, Faculty of Agriculture, Atatürk University, 25240, Erzurum, Turkey.

Authors' contributions

This work was carried out in collaboration between all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/BJAST/2016/27740

Editor(s):

(1) Ming-Chih Shih, Department of Health and Nutrition Science, Chinese Culture University, Taiwan.

Reviewers:

(1) Yiannis Kourkoutas, Democritus University of Thrace, Greece.

(2) Marcelo Teixeira Leite, Federal University of Paraíba, Brazil.

(3) Falodun Olutayo Israel, University of Ibadan, Ibadan, Nigeria.

Complete Peer review History: <http://www.sciencedomain.org/review-history/15690>

Mini-review Article

Received 16th June 2016
Accepted 27th July 2016
Published 6th August 2016

ABSTRACT

Flavour is an essential quality of food and formed by large number of volatile (alcohol, ketones, aldehydes, esters and terpenes etc.) and non-volatile compounds (amino acids and peptides etc.). In fermented sausage, flavour develops from biochemical reactions and is influenced by several variables such as formulation, process conditions and starter culture. Microorganisms involved in fermentation play an important role in the formation of aroma. Lactic acid bacteria and catalase-positive cocci are the most important starter cultures for fermented sausage. Lactic acid bacteria are mainly responsible for lactic acid production in sausage. Catalase-positive cocci have an impact on catalase activity, colour stability, prevention of rancidity and flavour formation. These microorganisms enhance flavour through proteolytic and lipolytic activity. This study reviews the functions of catalase-positive cocci and their effects on flavour in fermented sausages.

Keywords: *Catalase-positive cocci; fermented sausage; flavour; branched chained amino acid; volatile compounds.*

*Corresponding author: E-mail: abereketoglu@bayburt.edu.tr;

1. INTRODUCTION

Fermentation is one of the oldest methods for extending the shelf life of food products. Biochemical and physical reactions that take place during fermentation change the product's initial properties. Fermented sausages are produced from a mixture of meat, fat, curing agents (nitrate/nitrite), spices and sugars through lactic acid fermentation [1]. Lactic acid bacteria and catalase-positive cocci are the most important microorganisms in the technological and sensorial properties of dry or semi-dry fermented sausages [2-5]. Lactic acid bacteria play a major role in acidification [6]. Therefore, they improve the product's safety and stability. Gram-positive catalase-positive cocci play an important role in colour stability, prevention of rancidity and flavour formation [7-9].

Catalase-positive cocci grow slowly or not at all during ripening, and the growth occurs mainly on the surface of unsmoked sausages. Because of this, high levels of catalase-positive cocci (10^6 – 10^7 cfu/g) are added to sausage mixture. At this point, the specific enzyme activity of the strain is particularly important [10]. Slow ripening in fermented meat products provides an advantageous environment for catalase-positive cocci. These microorganisms are generally active until lactic acid formation results in low pH levels [11]. Adding small amounts of sugar at low ripening temperature causes slow acid formation, which encourages the growth of these microorganisms. Moreover, in fermented sausages produced using nitrate and low levels

of sugar, such as traditional Italian fermented sausages, catalase-positive cocci may be the dominant flora [12].

The typical flavour of dry fermented sausages is the result of a careful balance between volatile (alcohol, ketones, aldehydes, esters, terpenes, aliphatic hydrocarbons, aromatic hydrocarbons and furans) and non-volatile compounds (amino acids, peptides, sugars and nucleotides), which come from the basic ingredients (meat, spices, nitrites and other additives) or generated by the breakdown of carbohydrates, lipids and proteins through the action of microbial and endogenous meat enzymes during ripening [13]. This study reviews the functions of catalase-positive cocci and their effects on flavour in fermented sausages.

2. THE ROLE OF CATALASE-POSITIVE COCCI IN FERMENTED SAUSAGES

The catalase-positive cocci identified from various types of traditional fermented sausage are given in Table 1. *Staphylococcus xylosus* is the most frequently isolated catalase-positive cocci in dry fermented sausages [14-16]. Strains of *Kocuria* and coagulase-negative *Staphylococcus* are used as starter cultures. These species grow in the presence of 10% NaCl, produce catalase and reduce nitrate to nitrite. However, they exhibit slow growth under anaerobic conditions. In sausages which are ripened normally and slowly, there are fewer catalase-positive cocci in the core than the periphery after a few days of ripening, due to

Table 1. Catalase positive cocci strains identified from different traditional fermented sausages

Strains	Fermented sausage	References
<i>Staphylococcus xylosus</i>	Fermented Italian sausages	[17-20]
	Chorizo	[21]
	Slovak fermented sausage	[22]
	Fermented sausage (Basilicata region)	[23]
	Greek fermented sausage	[24,25]
	Sucuk (Turkish fermented sausage)	[16]
	Salami (Italian fermented sausages)	[26]
<i>S. saprophyticus</i>	Naples type salami	[27]
	Iberian type sausage	[28]
	Sucuk	[16]
<i>S. carnosus</i>	Artisanal Argentinean sausage	[29]
	Spanish dry-cured sausage	[30]
<i>S. succinus</i>	Sucuk	[16]
	Salami	[31]
<i>S. equorum</i>	Sucuk	[16]

oxygen sensitivity. *K. varians* in particular needs oxygen and it is less competitive in fermented sausage mixture in comparison to *S. carnosus* [10].

Catalase-positive cocci play an important role in product colour, stabilization, the prevention of rancidity and the emergence of volatile compounds that are effective in flavour. Effects of catalase positive cocci as starter culture on fermented meat products are given in Fig. 1. They are effective in the formation of nitrosomyoglobin, the compound responsible for the characteristic red colour of fermented sausages, by reducing nitrate into nitrite through nitrate reductase activity. Catalase-positive cocci utilise available free oxygen in the sausage after filling and provide necessary reductive conditions for colour stability. Catalase activity of these cocci decomposes hydrogen peroxide and thus the negative effects of peroxide on the colour of cured meat products are avoided [32,13]. In the case of high amounts of O₂ and insufficient enzyme activity for peroxide decomposition, H₂O₂ causes colour defects and early rancidity [13]. Catalase and superoxide dismutases of catalase-positive cocci significantly affect the sensorial properties of fermented sausage through antioxidant activity on lipid oxidation [33,34].

Nitrate reductase is an intracellular enzyme located on the cytoplasmic membrane. This enzyme reduces nitrate to nitrite. However, rapid drops in pH prevent microbial nitrate reduction and colour defects occur as a consequence. Thus, it is reported that the pH should not be decreased below 5.4 until a sufficient amount of nitrate has reduced to nitrite [13]. Several strains of staphylococci such as *S. carnosus*, *S. xylosus*, *S. equorum* and *S. lentus* have nitrate reductase activity [21,34-40].

The development of flavour is influenced by several variables such as product formulations, processing conditions, and starter cultures [13]. Olesen et al. [41] reported that sausages cured with nitrate have higher volatile compounds derived from lipid autooxidation, such as 1-pentanol, 1-hexanol and 2-heptanone, than sausages cured with nitrite. These differences are caused by antioxidative properties of nitrite. Stahnke [42] showed that glucose levels affect the amount of volatile compounds when *S. carnosus* and *S. xylosus* are used as starter cultures. In a study on *S. xylosus* and *S. carnosus* the amount of diacetyl and acetoin increased with glucose content, but ketones and

sulphides level decreased for *S. carnosus*. On the contrary, glucose blocks tricarboxylic acid cycle and has negative effects on the growth rate of *S. carnosus* [42] and *S. xylosus* [43].

Staphylococcus and *Kocuria* contribute to the development of colour and flavour by decomposing free amino acids and inhibiting the oxidation of unsaturated free fatty acids [44]. Different species of *Staphylococcus* produce different aroma compounds in different amounts [6,45]. The volatile compounds currently recognized as products of staphylococci are primarily amino acid catabolites, pyruvate metabolites, and methyl ketones from β -oxidation of fatty acids [7,9,46].

During meat fermentation, several volatile compounds are generated by the action of endogenous meat enzymes as well as the proteolytic and lipolytic activity of catalase-positive cocci. Many researchers have focused on proteolytic and lipolytic activity of catalase-positive cocci in sausage fermentation [15,40,47-50]. Because of their effects on colour, texture and flavour, catalase-positive cocci are commercially used as starter cultures with lactic acid bacteria to ferment meat. Among catalase-positive cocci, *S. xylosus* and *S. carnosus* are commonly used as starter cultures. *S. xylosus* and *S. carnosus* have several enzymes that contribute to protecting fermented sausages from the detrimental effects of oxygen. The antioxidative properties of *Staphylococcus* strains are responsible for a decrease in levels of volatile compounds derived from lipid oxidation reactions and prevent a reduction in sausage quality [33,51]. Also, the enzymes involved in the β -oxidation of fatty acids, which enhance the cured aroma in fermented sausages by increasing the concentration of methyl ketones, are purified from *S. Carnosus* [51]. On the other hand, *Staphylococcus* strains used as starter cultures have lipolytic activity [52], but it is also argued that lipase activity is limited under fermented sausage conditions [51].

For flavour development, proteolysis is the most important biochemical change. It involves the generation of amino acids, peptides, amines and aldehydes, which are precursors of volatile compounds [53-55]. It has been reported that *S. xylosus*, *S. equorum*, *S. carnosus*, *S. simulans*, *S. warneri* and *S. cohnii* strains that were isolated from dry cured meat products had proteolytic activity on sarcoplasmic and/or myofibrillar proteins [40,53,56].

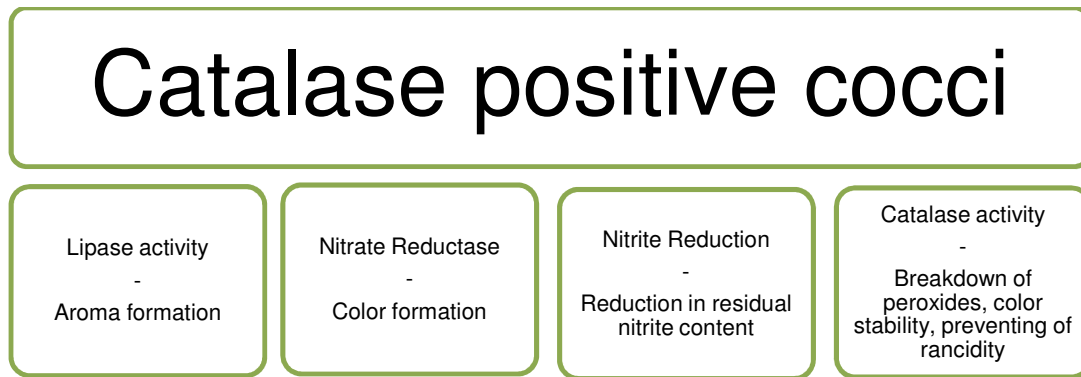


Fig. 1. Effects of catalase positive cocci used as starter culture on fermented meat products [8]

Aroma compounds originating from the degradation of amino acids are important for sausage flavour, especially the breakdown of leucine, isoleucine and valine into branched-chain methyl aldehydes and acids [7,57]. The initial step is the deamination of leucine catalyzed by branched-chain acid transferases, amino acid oxidases and dehydrogenases [58]. In one study, Stahnke [42] showed that the cell counts of *S. carnosus* and *S. xylosus* were correlated with methyl-branched aldehydes and ketones. Many factors can influence amino acid metabolism in these strains. Adding free amino acids to fermentation media did not affect total transaminase activity or volatile compounds. However, adding α -ketoglutarate increased the levels of methyl-branched alcohols, aldehydes, ethanol, 2-phenyl ethanol, phenylacetaldehyde, benzaldehyde, diacetyl, 4-methyl, 2-pentanone and ethyl esters. α -ketoglutarate increased transaminase activity. The limiting factor in the transamination of branched-chain amino acids (BCAAs) was the amount of amino group acceptors [59].

The decomposition of BCAAs leucine, valine and isoleucine into methyl-branched aldehydes, acids and alcohols by staphylococci strains has been studied by Masson et al. [60] Larroure et al. [61], Beck et al. [62], Olesen and Stahnke [63] and Olesen et al. [64]. It was shown that *S. carnosus* [6,7,60] and *S. xylosus* [62] have an effect on branched aldehydes (3-methyl butanal, 2-methyl butanal) and acids (3-methyl butanoic acid and 2-methyl butanoic acid) from amino acid degradation.

Leucine catabolism of *S. xylosus* [64] and *S. carnosus* [60, 61] were inhibited by nitrate. The formation of 3-methyl butanoic acid was significantly reduced by the presence of nitrate [60,61,64]. However, Stahnke [42] showed that

nitrate influences the production of 3-methyl butanoic acid from leucine catabolism. Another study reported that nitrite content increased the level of free amino acids while nitrate decreased in dry fermented sausages [65]. In contrast, Olesen et al. [64] determined that nitrite decreased the levels of volatile compounds through BCAA degradation.

The volatile profile of dry-fermented sausage consists of a wide variety of compounds such as aromatic and aliphatic hydrocarbons, aldehydes, alcohols, esters, sulphur containing compounds and furanes, etc. [6,48,66-68] and also most of those volatile compounds effect sausage flavour [57]. Aroma compounds generated from the degradation of amino acids play a major role in sausage flavour [42], especially the degradation of BCAAs into branched-chain alcohols, aldehydes, acids and esters [61,62].

It is thought that there is a correlation between amino acid and volatile compounds profiles. Waade and Stahnke [65] showed that amounts of valine, isoleucine and leucine are inversely correlated with 2-methyl propanal, 2-methyl butanal and 3-methyl butanal, respectively.

Masson et al. [60] showed that the use of nitrate has a strong effect on the formation of 3-methyl butanal in the presence of *S. carnosus*. However, it was also reported that adding nitrate decreased 3-methyl butanol and 3-methyl butanoic acid formation in the same strain. In another study on *S. xylosus* as a starter culture, nitrite reduced the formation of 3-methyl butanal and 2-methyl propionic acid by inhibiting amino acid degradation [48].

Olesen et al. [41] indicated that sausages with added nitrate have higher volatile compounds that originate from BCAA degradation than

sausages with added nitrite. That may be caused by the metabolic activity of the staphylococci strains that were used. The inhibition effect of nitrite on the degradation of BCAAs was also reported by Demeyer et al. [69] and Olesen et al. [64].

Some studies reported a correlation between catalase-positive cocci and acetoin formation. Montel et al. [7] characterized *S. saprophyticus* strains by their acetoin production capacity. Ravyts et al. [70] observed *S. sciuri*, *S. succinus* and *S. xylosus* strains as acetoin producers in southern European sausage.

In fermented sausages, another factor for aroma formation is the rate of fermentation. Researchers have reached different conclusions. According to the results of Tjener et al. [71] rapidly-acidified sausages showed high levels of ketones, sulphides and methyl-branched acids. However, there were high levels of methyl-branched alcohols, aldehydes and esters, methional and phenylacetaldehyde in slowly-acidified sausages in the same study. Ravyts et al. [70] showed that the acidification rate was depended on the catalase-positive cocci species. They reported that due to the survival rate of catalase-positive cocci in fermentation conditions, there is a volatile profile.

3. CONCLUSION

Many factors affect the volatile compounds generated by catalase-positive cocci in sausage fermentation. Studies strongly agree that volatile compounds result from the degradation of amino acid products by catalase-positive cocci. Flavour can be enhanced by regulating enzyme activities. Further studies on optimizing enzyme activity of catalase-positive cocci would lead to a better understanding of flavour formation during sausage fermentation.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Caplice E, Fitzgerald GF. Food fermentations: Role of microorganisms in food production and preservation. *International Journal of Food Microbiology*. 1999;50:131-49.
2. Aymerich T, Martín B, Garriga M, Hugas M. Microbial quality and direct PCR identification of lactic acid bacteria and nonpathogenic staphylococci from artisanal low-acid sausages. *Applied and Environmental Microbiology*. 2003;69(8): 4583-94.
3. Ammor S, Rachman C, Chaillou S, Prévost H, Dousset X, Zagorec M, et al. Phenotypic and genotypic identification of lactic acid bacteria isolated from a small-scale facility producing traditional dry sausages. *Food Microbiology*. 2005;22: 373-82.
4. Ammor MS, Mayo B. Selection criteria for lactic acid bacteria to be used as functional starter cultures in dry sausage production: An update. *Meat Science*. 2007;76:138-46.
5. Drosinos EH, Paramithiotis S, Kolovos G, Tsikouras I, Metaxopoulos I. Phenotypic and technological diversity of lactic acid bacteria and staphylococci isolated from traditionally fermented sausages in southern Greece. *Food Microbiology*. 2007;24:260-70.
6. Berdagué JL, Monteil P, Montel MC, Talon R. Effects of starter cultures on the formation of flavour compounds in dry sausage. *Meat Science*. 1993;35:275-87.
7. Montel MC, Reitz J, Talon R, Berdagué JL, Rousset-Akrim S. Biochemical activities of Micrococcaceae and their effects on the aromatic profiles and odours of a dry sausage model. *Food Microbiology*. 1996; 13:489-99.
8. Lücke FK. Fermented sausages, in: Wood BJB (Ed.). *Microbiology of fermented foods*. Blackie Academic and Professional, London; 1998.
9. Tjener K, Stahnke LH, Andersen L, Martinussen J. Growth and production of volatiles by *Staphylococcus carnosus* in dry sausages: Influence of inoculation level and ripening time. *Meat Science*. 2004;67: 447-52.
10. Lücke FK, Hechelmann H. Starterkulturen für Rohwurst und Rohschinken-zusammensetzung und Wirkung. *Mikrobiologie und Qualität von Rohwurst und Rohschinken*. Bundesanstalt Für Fleischforschung. 1985;193-218.
11. Hospital XF, Carballo J, Fernandez M, Arnau J, Gratacos M, Hierro E. Technological implications of reducing nitrate and nitrite levels in dry-fermented sausages: Typical microbiota, residual nitrate and nitrite and volatile profile. *Food Control*. 2015;57:275-81.
12. Lücke FK. Mikrobiologische Vorgänge bei der Herstellung von Rohwurst und

- rohschinken. mikrobiologie und qualität von rohwurst und rohschinken. Bundesanstalt Für Fleischforschung. 1985;85-102.
13. Kaban G, Kaya M, Lücke FK. Meat starter cultures. Encyclopedia of Biotechnology in Agriculture and Food. Taylor and Francis, Newyork; 2012.
 14. Comi G, Citterio B, Manzano M, Cantoni C, Bertoldi M. Evaluation and characterisation of Micrococcaceae strains in italian dry fermented sausages. Fleischwirtsch. 1992;72:1679-83.
 15. Miralles MC, Flores J, Perez-Martinez G. Biochemical tests for the selection of staphylococcus strains as potential meat starter cultures safety of raw meat fermented products. Food Microbiology. 1996;13:227–236.
 16. Kaban G, Kaya M. Identification of lactic acid bacteria and Gram-positive catalase-positive cocci isolated from naturally fermented sausage (sucuk). Journal of Food Science. 2008;73(8):385-88.
 17. Cocolin L, Manzano M, Aggio D, Cantoni C, Comi G. A novel polymerase chain reaction (PCR)-denaturing gradient gel electrophoresis (DGGE) for the identification of Micrococcaceae strains involved in meat fermentations. Its application to naturally fermented Italian sausages. Meat Science. 2001;58:59–64.
 18. Iacumin L, Comi G, Cantoni C, Cocolin L. Ecology and dynamics of coagulase-negative cocci isolated from naturally fermented Italian sausages. Systematic and Applied Microbiology. 2006;29:480-486.
 19. Iacumin L, Manzano M, Comi G. Catalase-positive cocci in fermented sausage: Variability due to different pork breeds, breeding systems and sausage production technology. Food Microbiology. 2012;29: 178-186.
 20. Rebecchi A, Pisacane V, Callegari ML, Puglisi E, Morelli L. Ecology of antibiotic resistant coagulase-negative staphylococci isolated from the production chain of a typical Italian salami. Food Control. 2015; 53:14-22.
 21. García-Varona M, Santos EM, Jaime I, Rovira J. Characterisation of Micrococcaceae isolated from different varieties of chorizo. International Journal of food Microbiology. 2000;54:189-195.
 22. Simonová M, Stropfová V, Marciňáková M, Lauková A, Vesterlund S, Moratalla ML, et al. Characterization of *Staphylococcus xylosus* and *Staphylococcus carnosus* isolated from Slovak meat products. Meat Science. 2006;73:559–64.
 23. Bonomo MG, RicciardiZotta T, Sico M, Salzano G. Technological and safety characterization of coagulase-negative staphylococci from traditionally fermented sausages of Basilicata region (Southern Italy). Meat Science. 2009;83:15–23.
 24. Papamanoli E, Kotzekidou P, Tzanetakis N, Litopoulou- Tzanetaki E. Characterization of Micrococcaceae isolated from dry fermented sausage. Food Microbiology. 2002;19:441-49.
 25. Drosinos EH, Mataragas M, Xiraphi N, Moschonas G, Gaitis F, Metaxopoulos J. Characterization of the microbial flora from a traditional Greek fermented sausage. Meat Science. 2005;69:307-317.
 26. Greppi A, Ferrocino I, La Storia A, Rantsiou K, Ercolini D, Cocolin L. Monitoring of the microbiota of fermented sausages by culture independent rRNA-based approaches. International Journal of Food Microbiology. 2015;212:67-75.
 27. Coppola S, Mauriello G, Aponte M, Moschetti G, Villani F. Microbial succession during ripening of Naples-type salami, a southern Italian fermented sausage. Meat Science. 2000;56:321–29.
 28. Benito MJ, Serradilla MJ, Martín A, Aranda E, Hernández A, Córdoba MG. Differentiation of staphylococci from iberian dry fermented sausages by protein fingerprinting. Food Microbiology. 2008;25: 676–82.
 29. Fontana C, Cocconcelli PS, Vignolo G. Monitoring the bacterial population dynamics during fermentation of artisanal Argentinean sausages. International Journal of Food Microbiology. 2005; 103(2):131-142.
 30. Landeta G, Curiel J, Carrascosa V, Muñoz R, de las Rivas B. Characterization of coagulase-negative staphylococci isolated from Spanish dry cured meat products. Meat Science. 2013;93:387–96.
 31. Polka J, Rebecchi A, Pisacane V, Morelli L, Puglisi E. Bacterial diversity in typical Italian salami at different ripening stages as revealed by High-throughput sequencing of 16S rRNA amplicons. Food Microbiology. 2015;46:342-356.
 32. Kaya M, Kaban G. Fermented meat products, In: Aran N. (Ed), Food Biotechnology. Nobel Publishing, Ankara. 2010;157-90.

33. Barrière C, Leroy-Sétrin S, Talon R. Characterization of catalase and superoxide dismutase in *Staphylococcus carnosus* 833 strain. *Journal of Applied Microbiology*. 2001;91:514–19.
34. Mauriello G, Casaburi A, Blaiotta G, Villani F. Isolation and technological properties of coagulase negative staphylococci from fermented sausages of Southern Italy. *Meat Science*. 2004;67:149–158.
35. Neubauer H, Gotz F. Physiology and interaction of nitrate and nitrite reduction in *Staphylococcus carnosus*. *J. Bacteriol*. 1996;178:2005–9.
36. Coppola R, Iorizzo M, Saotta R, Sorrentino E, Grazia L. Characterization of micrococci and staphylococci isolated from soppressata molisana, a Southern Italy fermented sausage. *Food Microbiology*. 1997;14:47-53.
37. Pantel I, Lindgren PE, Neubauer H, Götz F. Identification and characterization of the *Staphylococcus carnosus* nitrate reductase operon. *Mol. Gen. Genet*. 1998;259:105-14.
38. Talon R, Walter D, Chartier S, Barrière C, Montel MC. Effect of nitrate and incubation conditions on the production of catalase and nitrate reductase by Staphylococci. *International Journal of Food Microbiology*. 1999;52:47–56.
39. Fedtke I, Kamps A, Krismer B, Götz F. The nitrate reductase and nitrite reductase operons and the *narT* gene of *Staphylococcus carnosus* are positively controlled by the novel Two-component system NreBC. *Journal of Bacteriology*. 2002;184(23):6624-34.
40. Casaburi A, Blaiotta G, Mauriello G, Pepe O, Villani F. Technological activities of *Staphylococcus carnosus* and *Staphylococcus simulans* strains isolated from fermented sausages. *Meat Science*. 2005;71:643–50.
41. Olesen PT, Meyer AS, Stahnke LH. Generation of flavour compounds in fermented sausages-the influence of curing ingredients, *Staphylococcus* starter culture and ripening time. *Meat Science*. 2004;66:675–87.
42. Stahnke LH. Volatiles produced by *Staphylococcus xylosus* and *Staphylococcus carnosus* during growth in sausage minces Part II. The Influence of Growth Parameters. *Lebensmittel-Wissenschaft & Technologie*. 1999;6:365–371.
43. Sørensen BB, Jakobsen M. The combined effects of environmental conditions related to meat fermentation on growth and lipase production by the starter culture *Staphylococcus xylosus*. *Food Microbiology*. 1996;13:265–74.
44. Talon R, Leroy S, Lebert I. Microbial ecosystems of traditional fermented meat products: The importance of indigenous starters. *Meat Science*. 2007;77:55–62
45. Kaban G, Kaya M. Effects of *Staphylococcus carnosus* on quality characteristics of sucuk (Turkish dry-fermented sausage) during ripening. *Food Science and Biotechnology*. 2009;18(1): 150-56.
46. Cocconcelli PS. Starter cultures: Bacteria, in: F. Toldra` (ed.), *Handbook of fermented meat and poultry*. Balckwell Publishing, Oxford; 2007.
47. Johansson G, Berdagué JL, Larsson M, Tran N, Borch E. Lipolysis, proteolysis and formation of volatile components during ripening of a fermented sausage with *Pediococcus pentosaceus* and *Staphylococcus xylosus* as starter cultures. *Meat Science*. 1994;38:203–18.
48. Stahnke LH. Dried sausages fermented with *Staphylococcus xylosus* at different temperatures and with different ingredient levels Part II. Volatile components. *Meat Science*. 1995;41:193–209.
49. Talon R, Montel MC. Hydrolysis of esters by staphylococci. *International Journal of food Microbiology*. 1997;36:207-14.
50. Engelvin G, Feron G, Perrin C, Molle D, Talon R. Identification of β -oxidation and thioesterase activities in *Staphylococcus carnosus* 833 strain. *FEMS Microbiology Letters*. 2000;190:115–20.
51. Flores M, Toldrá F. Microbial enzymatic activities for improved fermented meats. *Trends in Food Science & Technology*. 2011;22:81–90.
52. Kenneally PM, Leuschner RG, Arendt EK. Evaluation of the lipolytic activity of starter cultures for meat fermentation purposes. *Journal of Applied Microbiology*. 1998;84, 839–46.
53. Rodríguez M, Núñez F, Córdoba JJ, Bermúdez ME, Asensio MA. Evaluation of proteolytic activity of micro-organisms isolated from dry cured ham. *Journal of Applied Microbiology*. 1998;85:905–12.
54. Hughes MC, Kerry JP, Arendt EK, Kenneally PM, McSweeney PLH, O'Neill EE. Characterization of proteolysis during

- the ripening of semi-dry fermented sausages. *Meat Science*. 2002;62:205-16.
55. Roseiro LC, Santos C, Sol M, Borges MJ, Anjos M, Gonçalves H, Carvalho AS. Proteolysis in painho de portalegre dry fermented sausage in relation to ripening time and salt content. *Meat Science*. 2008; 79:784–94.
 56. Mauriello G, Casaburi A, Villani F. Proteolytic activity of *Staphylococcus xylosus* strains on pork myofibrillar and sarcoplasmic proteins and use of selected strains in the production of 'Naples type' salami. *Journal of Applied Microbiology*. 2002;92:482-90.
 57. Stahnke LH. Dried sausages fermented with *Staphylococcus xylosus* at different temperatures and with different ingredient levels Part III. Sensory evaluation. *Meat Science*. 1995;41:211–23.
 58. Massey LK, Sokatch JR, Conrad RS. Branched-chain amino acid catabolism in bacteria. *Bacteriological Reviews*. 1976;40: 42–54.
 59. Tjener K, Stahnke LH, Andersen L, Martinussen J. Addition of α -ketoglutarate enhances formation of volatiles by *Staphylococcus carnosus* during sausage fermentation. *Meat Science*. 2004;67:711-19.
 60. Masson F, Hinrichsen L, Talon R, Montel MC. Factors influencing leucine catabolism by a strain of *Staphylococcus carnosus*. *Food Microbiology*. 1999;49:173–78.
 61. Larrouture C, Ardaillon V, Pépin M, Montel M. Ability of meat starter cultures to catabolize leucine and evaluation of the degradation products by using an HPLC method. *Food Microbiology*. 2000;17:563–70.
 62. Beck HC, Hansen AM, Lauritsen FR. Metabolite production and kinetics of branched-chain aldehyde oxidation in *Staphylococcus xylosus*. 2002;31:94–101.
 63. Olesen PT, Stahnke LH. The influence of precultivation parameters on the catabolism of branched-chain amino acids by *Staphylococcus xylosus* and *Staphylococcus carnosus*. *Food Microbiology*. 2003;20:621–29.
 64. Olesen PT, Stahnke LH, Talon R. Effect of ascorbate, nitrate and nitrite on the amount of flavour compounds produced from leucine by *Staphylococcus xylosus* and *Staphylococcus carnosus*. *Meat Science*. 2004;68:193–200.
 65. Waade C, Stahnke LH. Dried sausages fermented with *Staphylococcus xylosus* at different temperatures and with different ingredient levels. Part IV. Amino acid profile. *Meat Science*. 1997;46:101–14.
 66. Schmidt S, Berger RG. Aroma compounds in fermented sausages of different origins. *LWT - Food Science and Technology*. 1998;31:559–67.
 67. Ansorena D, Gimeno O, Astiasaran I, Bello J. Analysis of volatile compounds by GC±MS of a dry fermented sausage: Chorizo de Pamplona. *Food Research International*. 2001;34:67-75.
 68. Kaban G, Kaya M. Effects of *Lactobacillus plantarum* and *Staphylococcus xylosus* on the quality characteristics of dry fermented sausage "sucuk". *Journal of Food Science*. 2009;74:58–63.
 69. Demeyer D, Raemaekers M, Rizzo A, Holck A, De Smedt A, ten Brink B et al.. Control of bioflavour and safety in fermented sausages: First results of a European project. *Food Research International*. 2000;33:171–80.
 70. Ravyts F, Steen L, Goemaere O, Paelinck H, De Vuyst L, Leroy F. The application of staphylococci with flavour-generating potential is affected by acidification in fermented dry sausages. *Food Microbiology*. 2010;27:945–54.
 71. Tjener K, Stahnke LH, Andersen L, Martinussen J. A fermented meat model system for studies of microbial aroma formation. *Meat Science*. 2004;66:211-18.

© 2016 Kamiloğlu et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://sciedomain.org/review-history/15690>